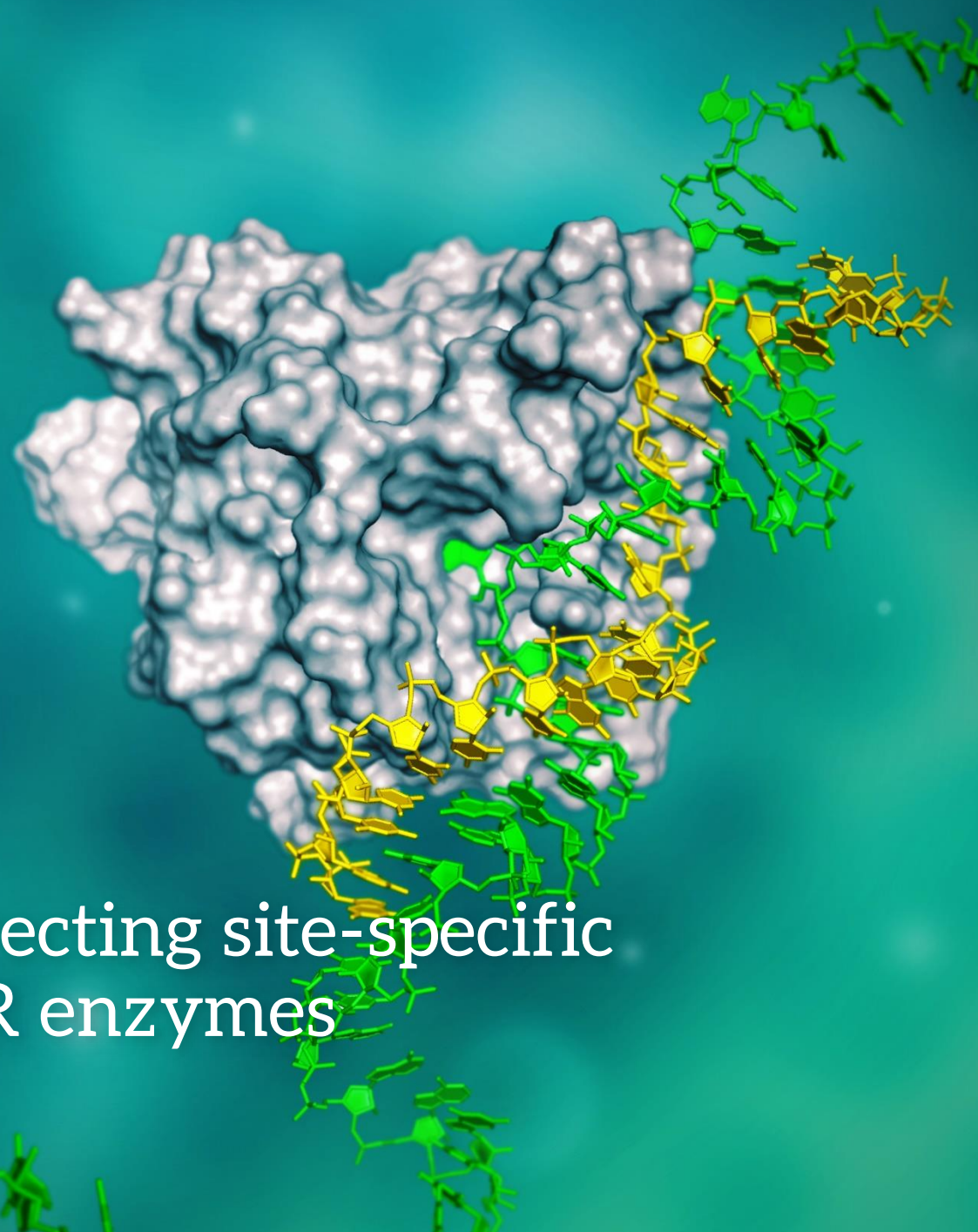




# AXIOMER<sup>®</sup> TECHNOLOGY

Therapeutic oligonucleotides for directing site-specific  
A-to-I editing by endogenous ADAR enzymes



# Forward looking statements

This presentation contains forward-looking statements that involve substantial risks and uncertainties. All statements, other than statements of historical facts, contained in this presentation, including but not limited to, statements regarding our strategy, future operations, future preclinical and clinical trial plans and related timing of trials and results, research and development, future financial position, future revenues, projected costs, prospects, therapeutic potential of our product candidates, plans and objectives of management, are forward-looking statements. The words “aim,” “anticipate,” “believe,” “estimate,” “expect,” “intend,” “may,” “plan,” “predict,” “project,” “target,” “potential,” “will,” “would,” “could,” “should,” “continue,” and similar expressions are intended to identify forward-looking statements, although not all forward-looking statements contain these identifying words.

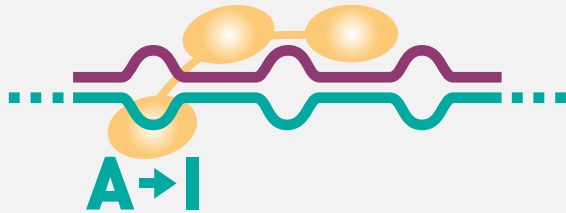
Forward-looking statements represent our management’s beliefs and assumptions only as of the date of this presentation. We may not actually achieve the plans, intentions or expectations

disclosed in our forward-looking statements, and you should not place undue reliance on our forward-looking statements. Actual results or events could differ materially from the plans, intentions and expectations disclosed in the forward-looking statements we make. The forward-looking statements contained in this presentation reflect our current views with respect to future events, and we assume no obligation to update any forward-looking statements except as required by applicable law. These forward-looking statements are subject to a number of risks, uncertainties and assumptions, including those that may be described in greater detail in the annual report filed on Form 20-F for the year ended December 31, 2018 that we have filed with the U.S. Securities and Exchange Commission (the “SEC”) and any subsequent filings we have made with the SEC. We have included important factors in the cautionary statements included in that annual report, particularly in the Risk Factors section, and subsequent filings with the SEC that we believe could cause actual results or events to differ materially from the forward-looking statements that we make.

# Therapeutic potential

# Axiomer RNA editing platform

*Editing Oligonucleotide (EON) mediated A-to-I editing*



## Unique A-to-I RNA editing

- A-to-I editing in RNA
- Using endogenous ADAR
- ADAR is recruited by a single stranded Editing OligoNucleotide (EON)
- I is translated as a G, allowing to target G-to-A mutations
- Specific, potent and stable by *design*



## Strong IP protection

- Invented in house at ProQR laboratories
- Foundational patents owned by ProQR
- Unrivalled know how on EON design and high-throughput assays
- Key collaborations with ADAR experts in the world

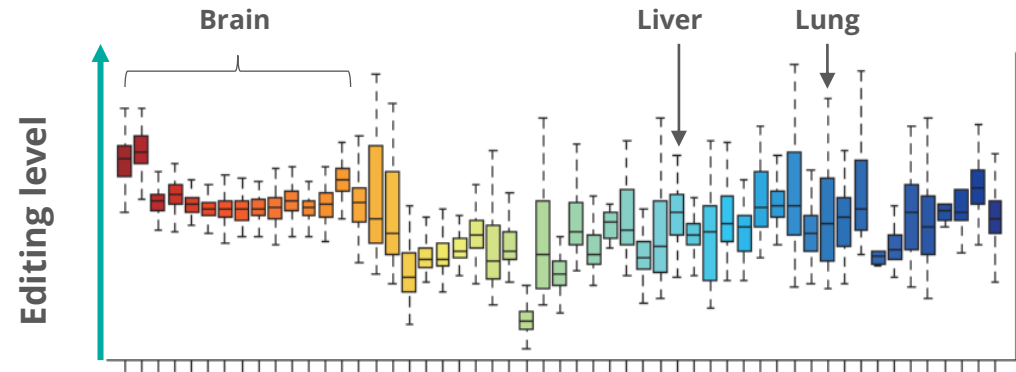
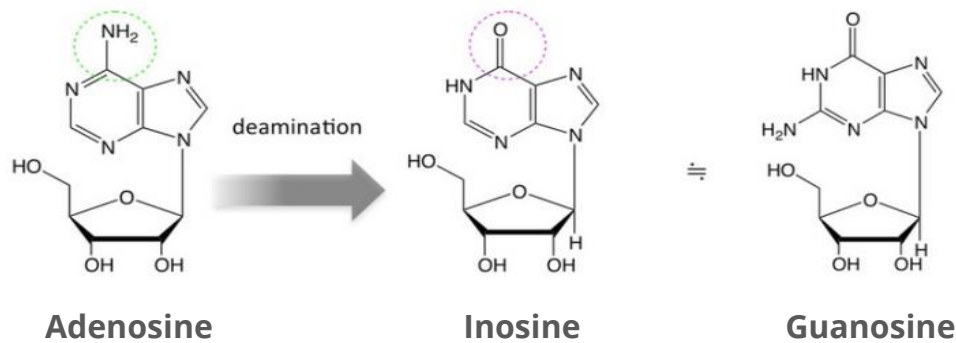


## Broad applicability

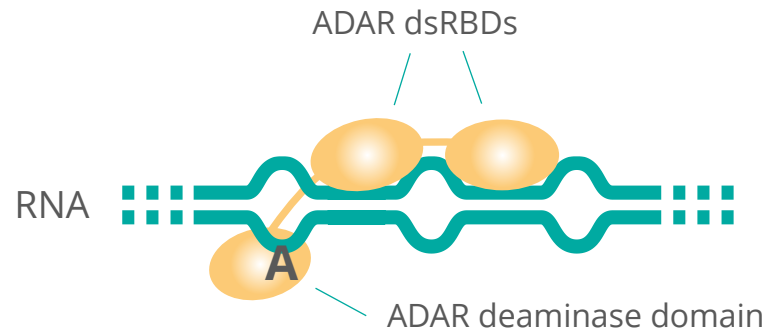
- Axiomer technology can target G-to-A mutations
- 22,000 G-to-A mutations causing disease
- Broader applicability in RNA and protein engineering for medical purposes

# A-to-I editing: Therapeutic opportunity

*The most prevalent editing event in human tissues*



Adapted from Tan et al. 2017 Nature 550:249-254

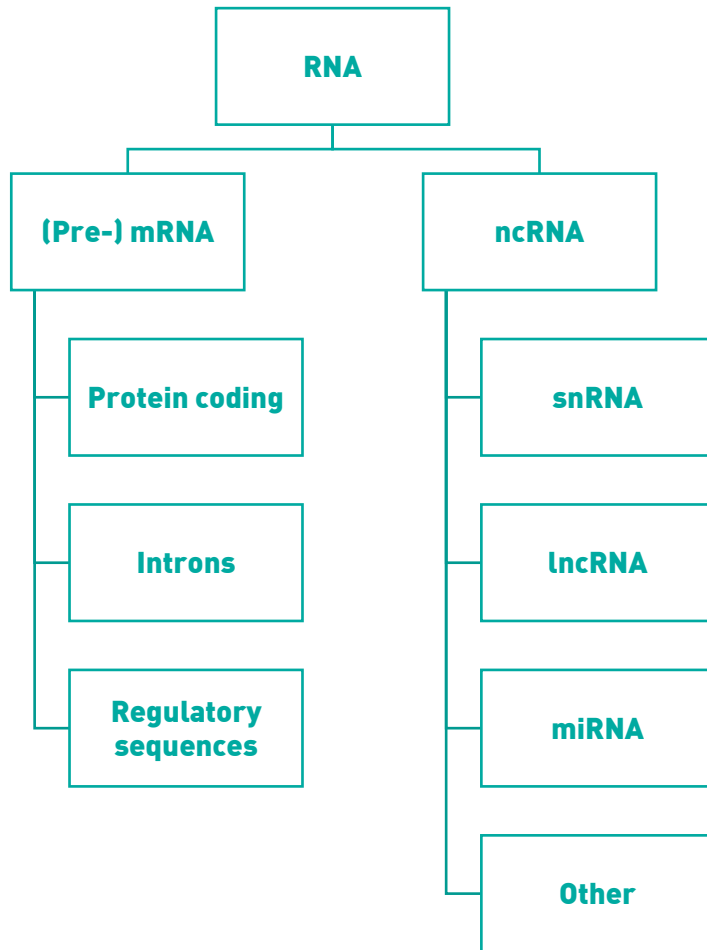


- No sequence dependence
- 4 million ADAR sites in the human transcriptome
- Extent of editing similar in most human tissues, making therapeutic editing feasible in all disease areas

- Human catalytic ADARs: ADAR1 and ADAR2
- A-to-I editing occurs in both nucleus and cytoplasm

# Axiomer<sup>®</sup> is widely applicable

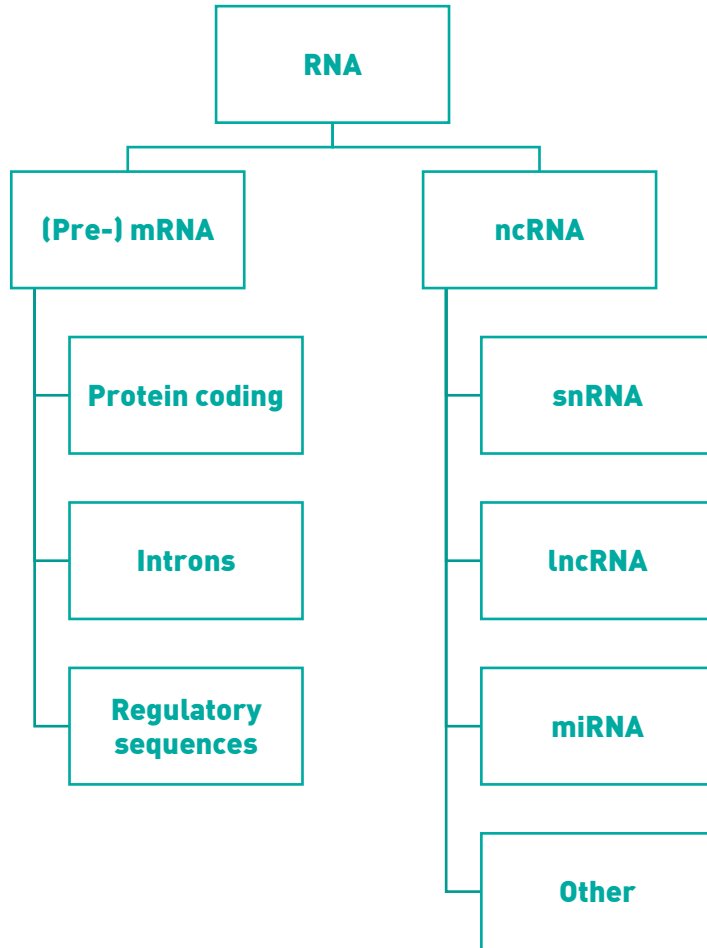
*Examples of different target RNAs*



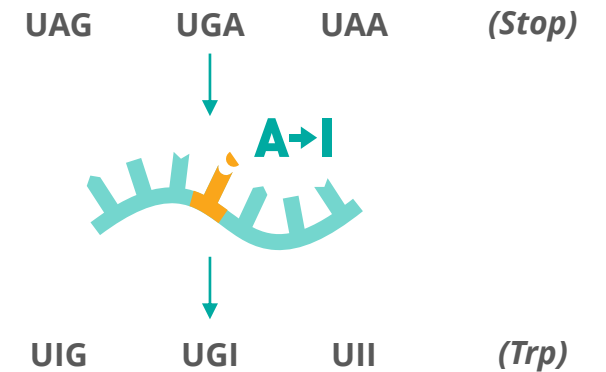
| Effect on RNA              | Effect of therapy                     |
|----------------------------|---------------------------------------|
| Correct nonsense mutations | Restore translation                   |
| Correct missense mutations | Restore protein                       |
| Correct splice sites       | Restore splicing of mRNA              |
| Modify codon               | Alter protein function                |
| Modify protein binding     | Alter expression                      |
| Modify 2° RNA structure    | Alter expression                      |
| Alter regulatory sequence  | Modify target RNA/DNA/protein binding |

# Axiomer<sup>®</sup> is widely applicable

Stop codons as PoC for a wide class of disease indications



| Effect on RNA              | Effect of therapy                     |
|----------------------------|---------------------------------------|
| Correct nonsense mutations | Restore translation                   |
| Correct missense mutations | Restore protein                       |
| Correct splice sites       | Restore splicing of mRNA              |
| Modify codon               | Alter protein function                |
| Modify protein binding     | Alter expression                      |
| Modify 2° RNA structure    | Alter expression                      |
| Alter regulatory sequence  | Modify target RNA/DNA/protein binding |



- Class of mutations resulting in complete loss of function
- First proof of concept for the Axiomer<sup>®</sup> approach

# Axiomer® EONs

Molecular basis for targeted A-to-I editing

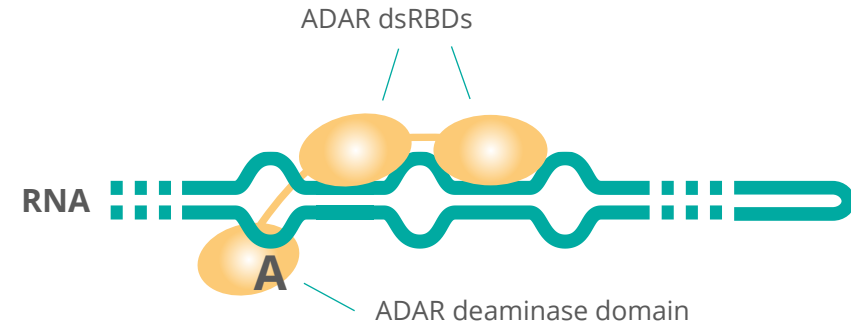
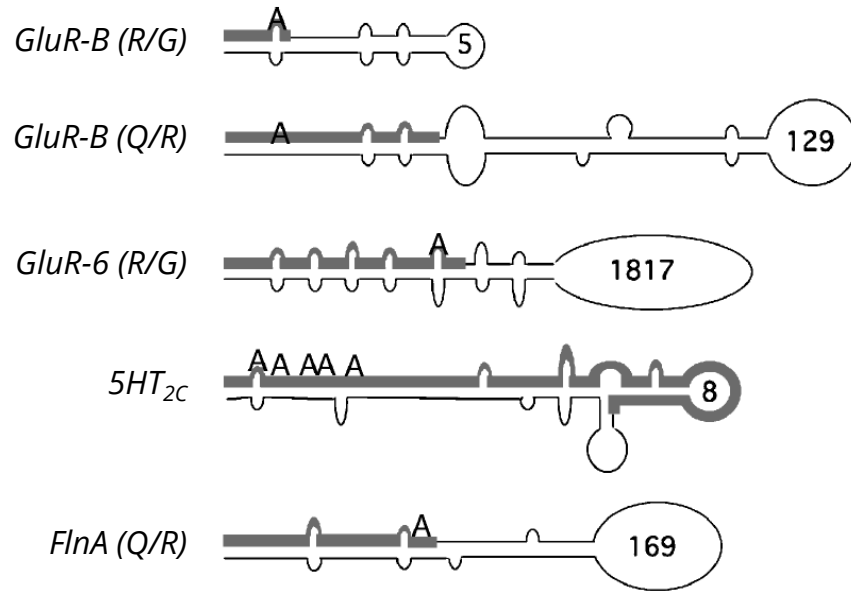


# ADARs deaminate adenosine in dsRNA

*Endogenous editing on natural substrates*

## ADAR targets:

Adenosines in dsRNAs with incomplete helices



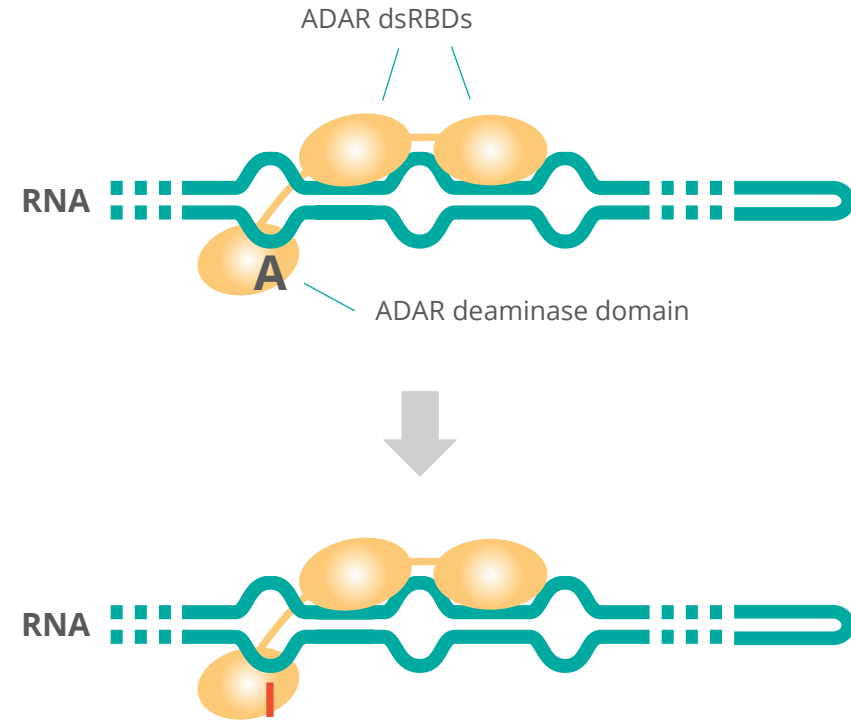
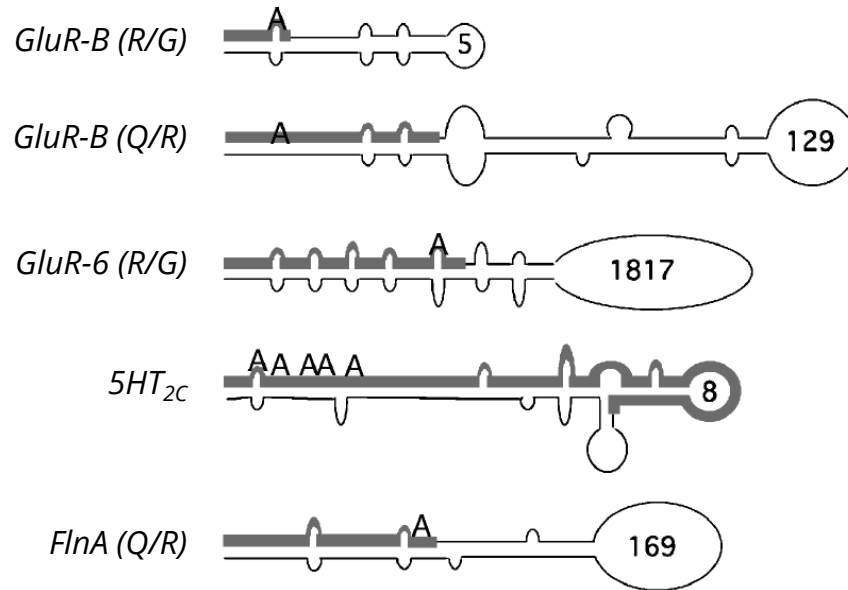
Wahlstedt & Öhman 2011, WIREs RNA

# ADARs deaminate adenosine in dsRNA

*Endogenous editing on natural substrates*

## ADAR targets:

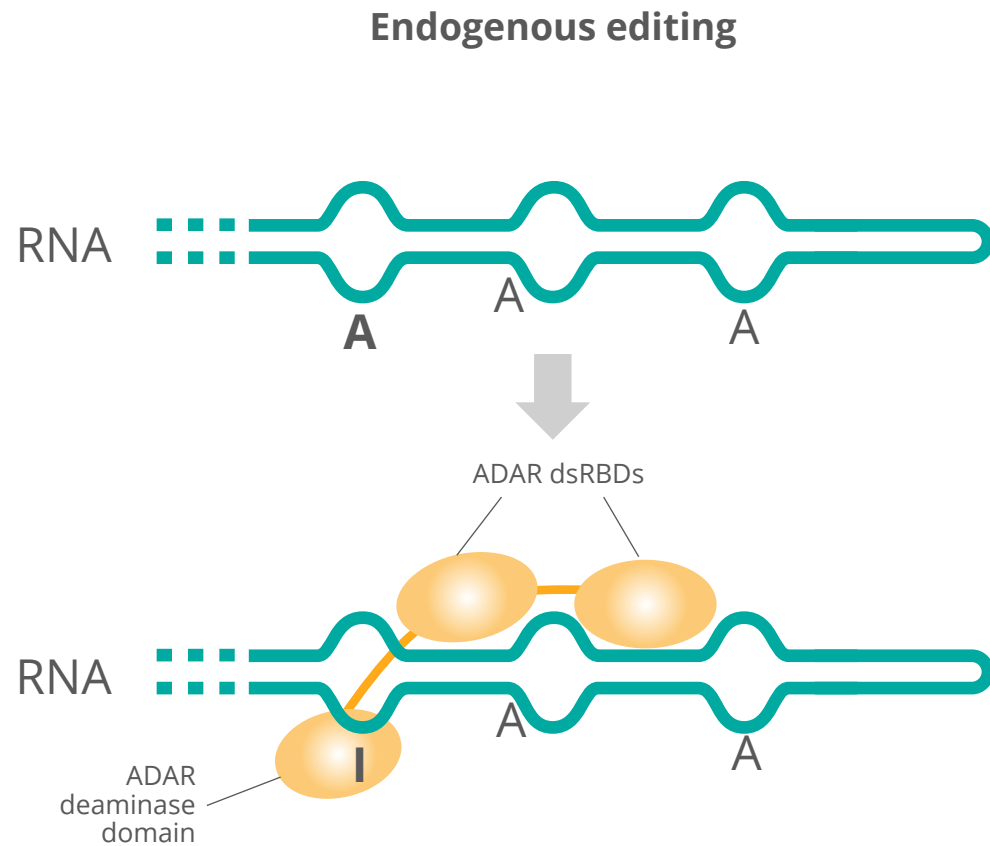
Adenosines in dsRNAs with incomplete helices



Wahlstedt & Öhman 2011, WIREs RNA

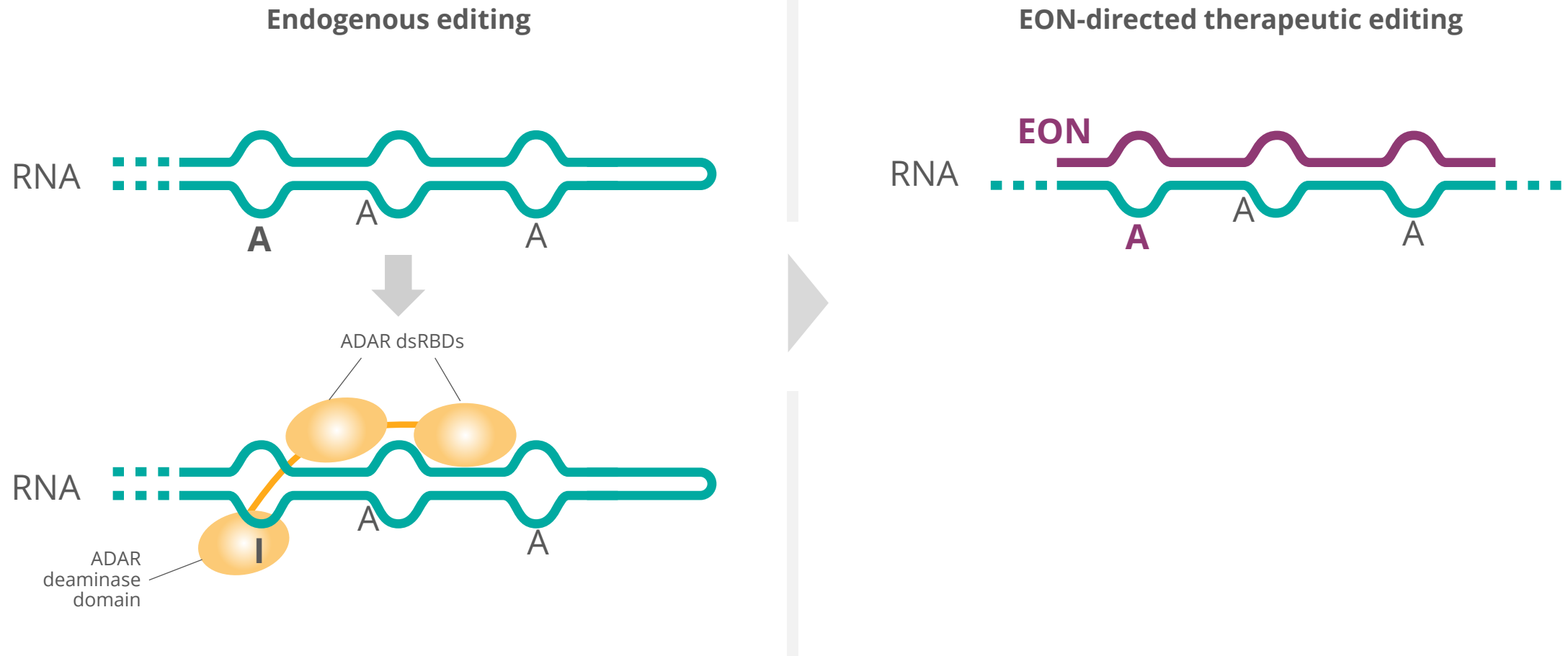
# EONs designed for targeted editing (1)

*Mimicking natural RNA editing*



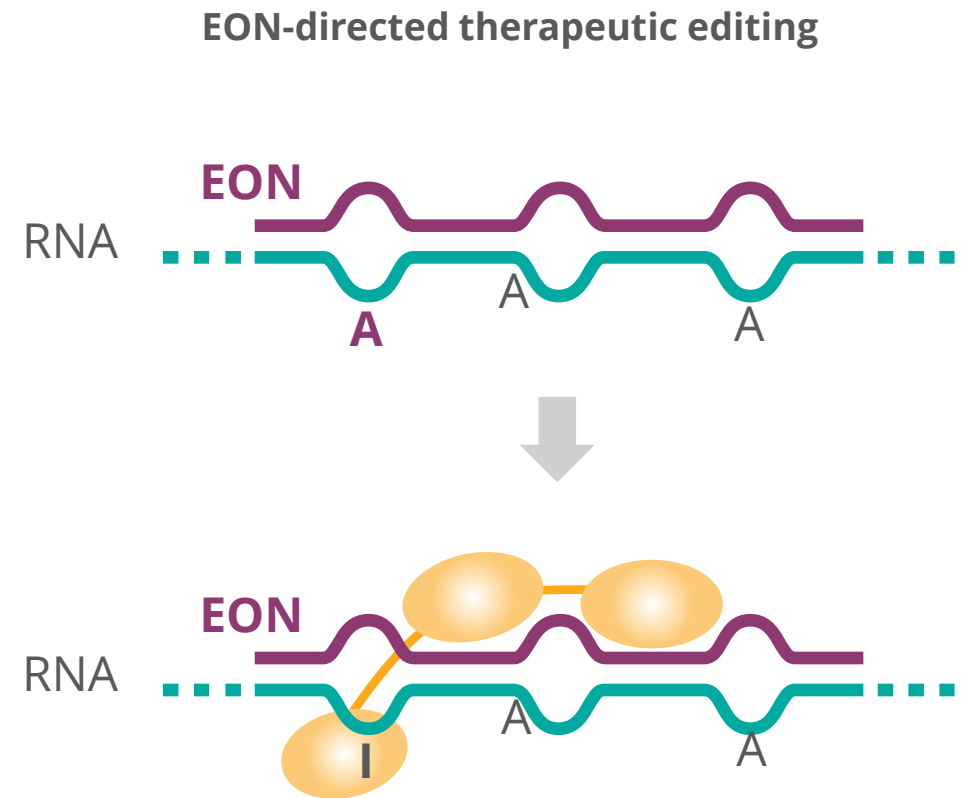
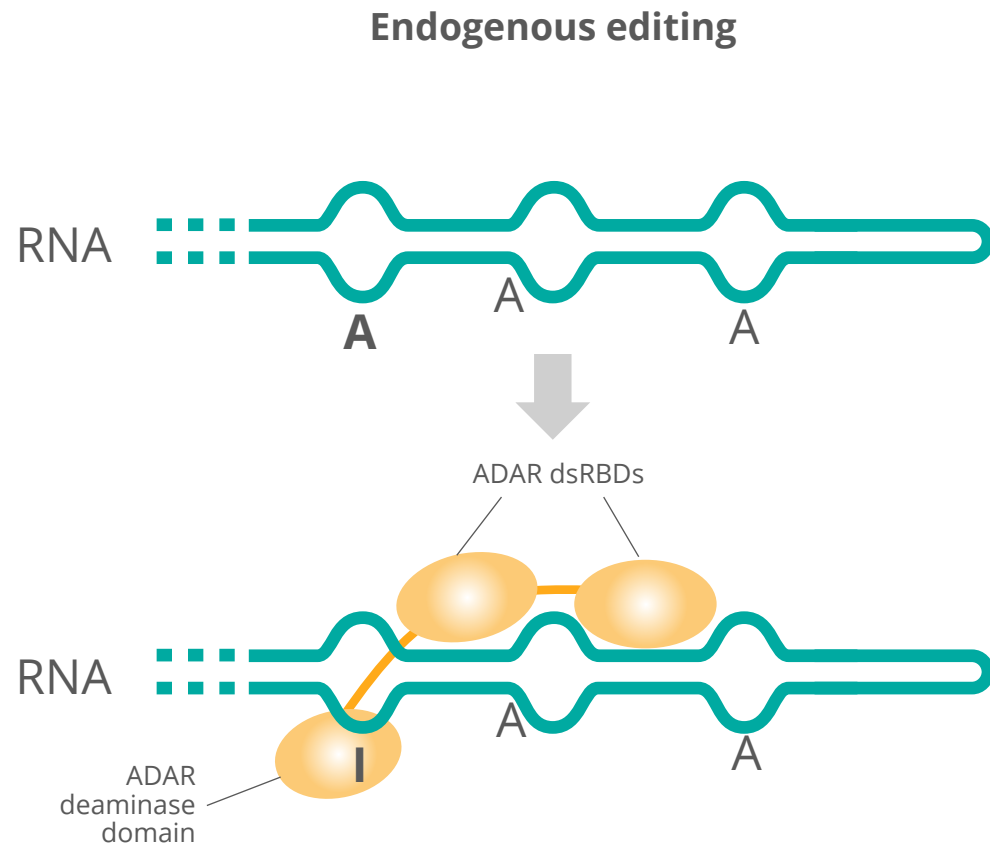
# EONs designed for targeted editing (2)

*EON and the target RNA form a double helix*



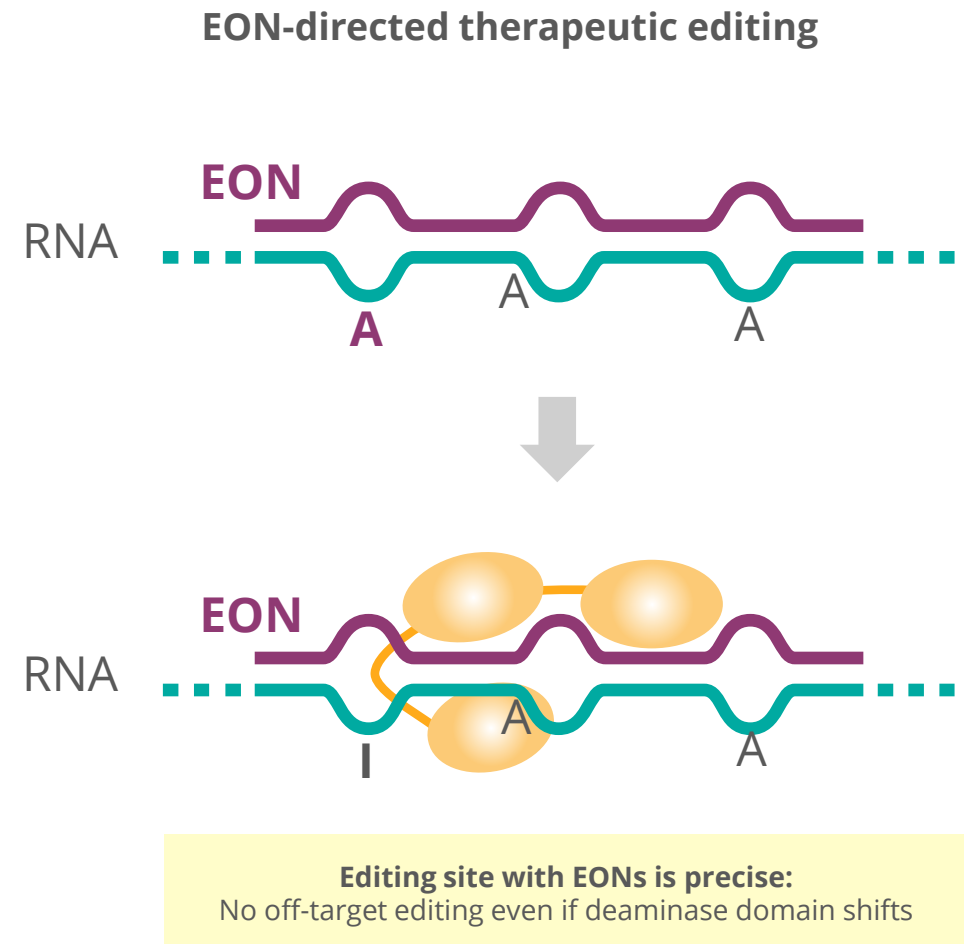
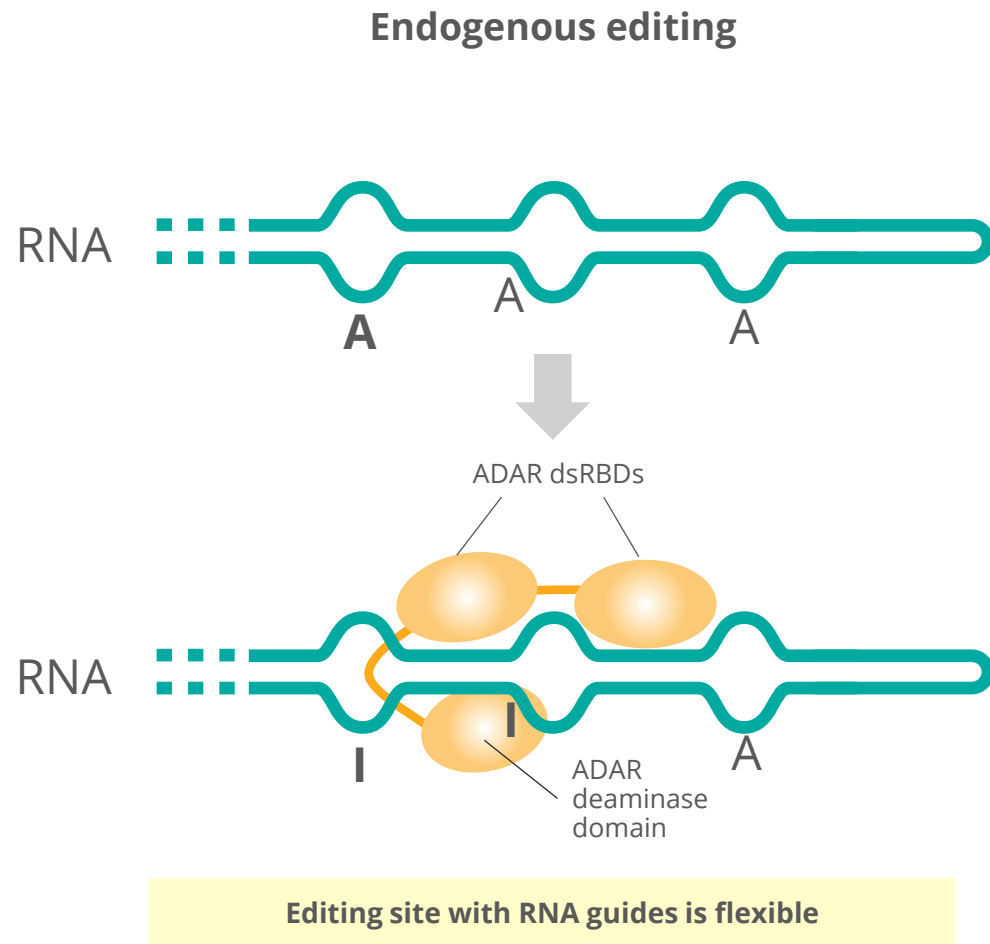
# EONs designed for targeted editing (3)

*ADAR deaminates the target A in EON-target RNA helix*



# EONs designed for targeted editing (4)

*Advantage over RNA guides: Specificity*



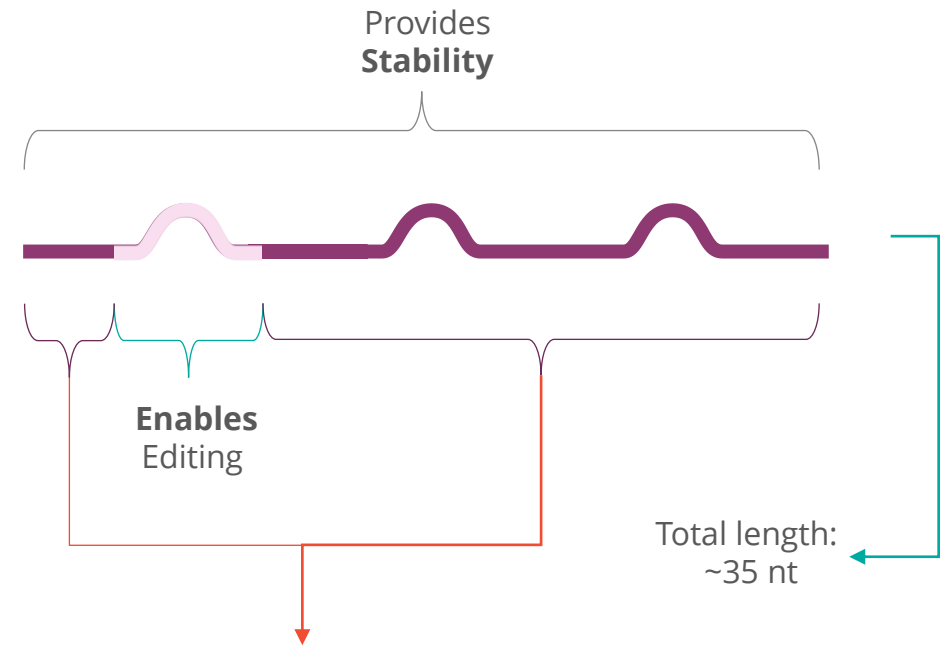
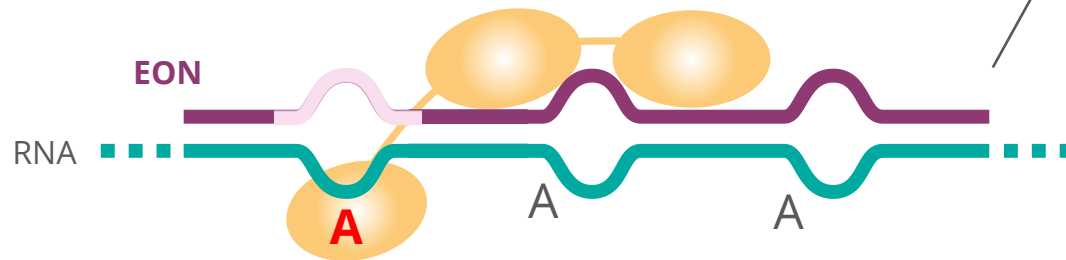
# EONs designed for targeted RNA editing

*Functionality defined by sequence and chemistry*

## Sequence defines target RNA binding

### EON chemical modifications define:

- Editing specificity
- Stability (nuclease resistance)
- Bioavailability
- Cell and tissue uptake



Backbone modifications enable ADAR binding, and **disable** off-target editing

# Structural basis for nt modifications

*ADAR binding and catalysis require different modifications*

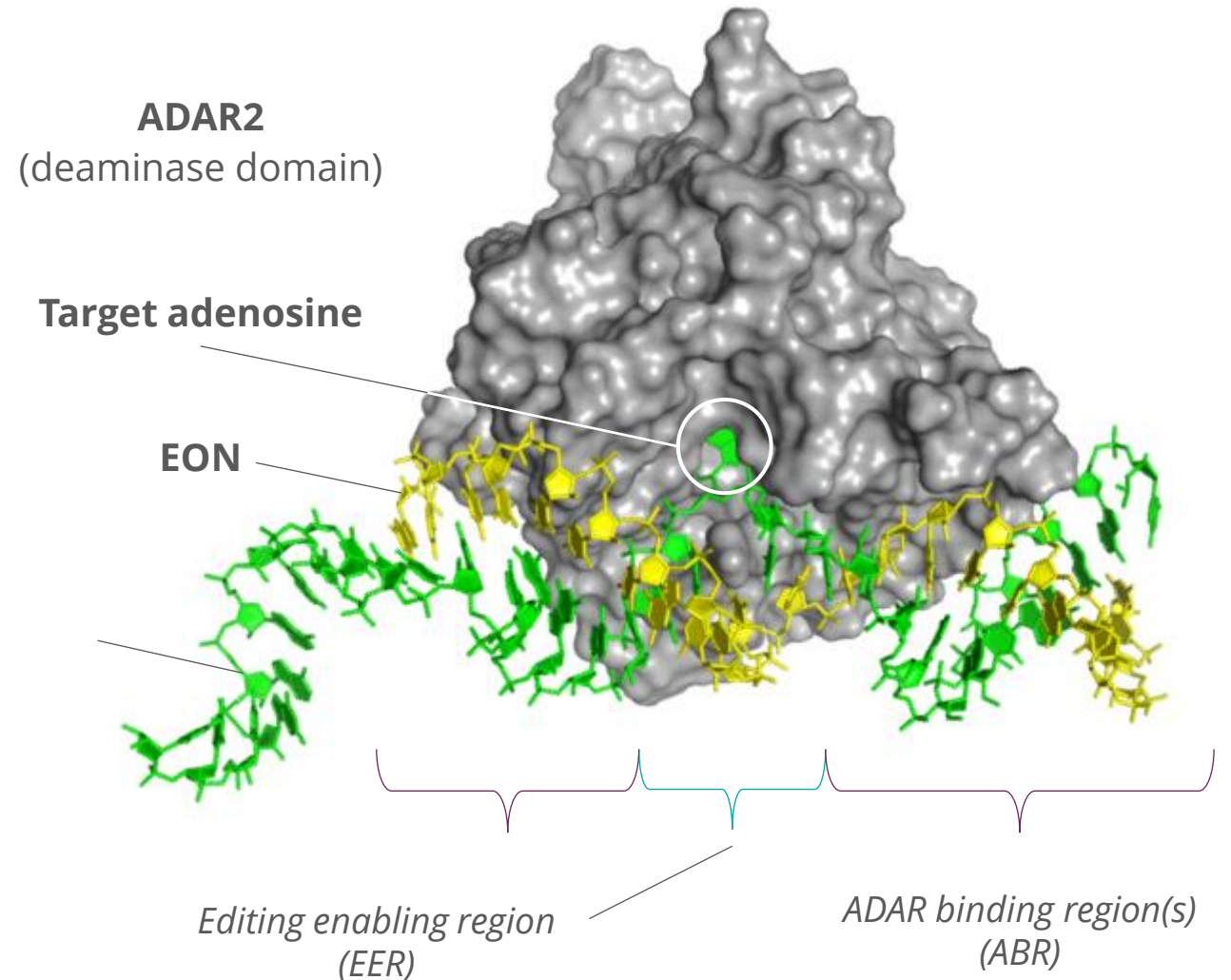
## ADAR binding region

- Modifications that are compatible with ADAR binding, but do not fit in the catalytic center

## Editing enabling region

- Modifications that fit into the catalytic center

Structural modelling provides a **basis for further optimization of EONs**





# Axiomer® EONs

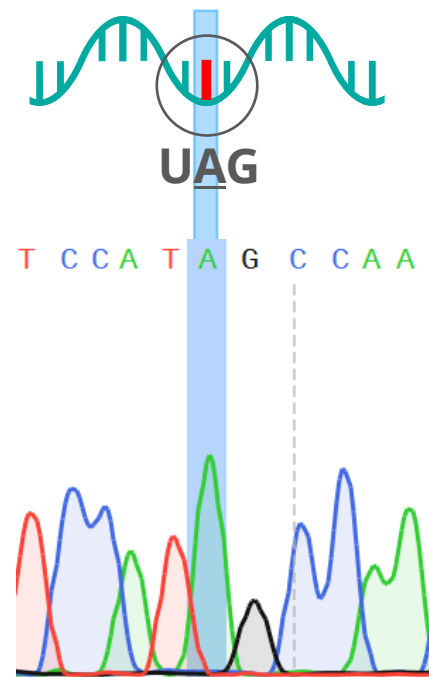
PoC studies for targeted editing

# EONs can restore ORFs

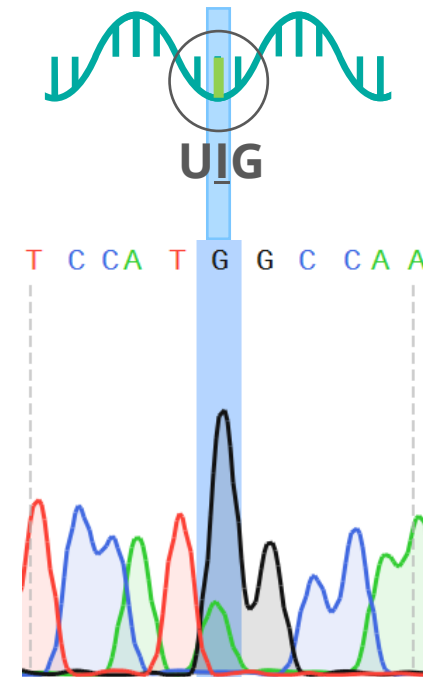
*In vitro proof of concept in a GFP reporter*

- GFP W57X reporter in Hepa1-6 cells
- ADAR2 overexpression
- Transfection with 100 nM EON
- Readout by Sanger sequencing of the RT-PCR product

W57X Mutation



Mutation treated with EON



85% of transcript corrected by editing

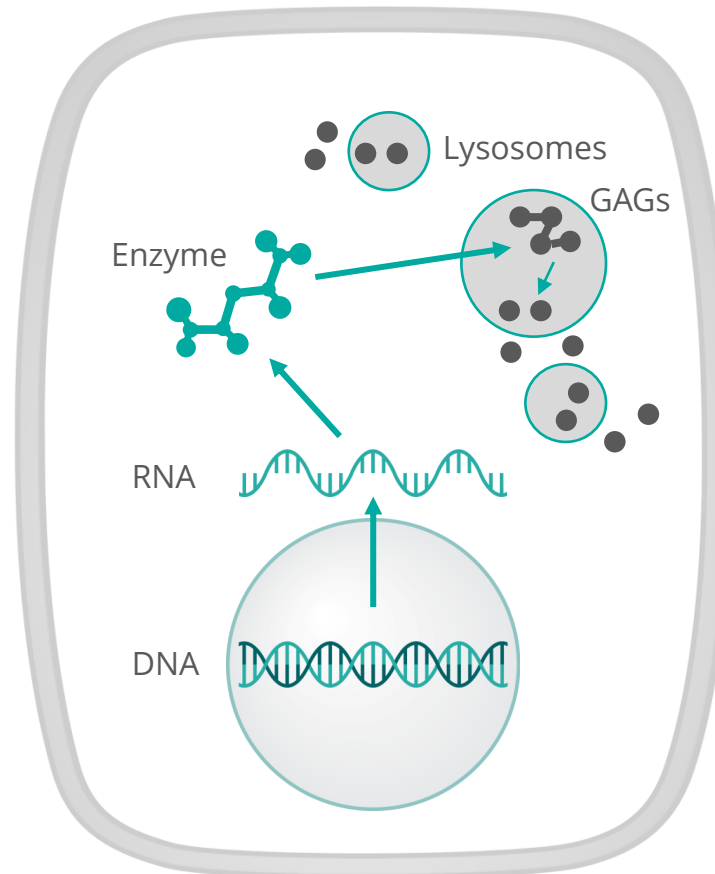
# Model system for *in vivo* PoC:

## Hurler syndrome

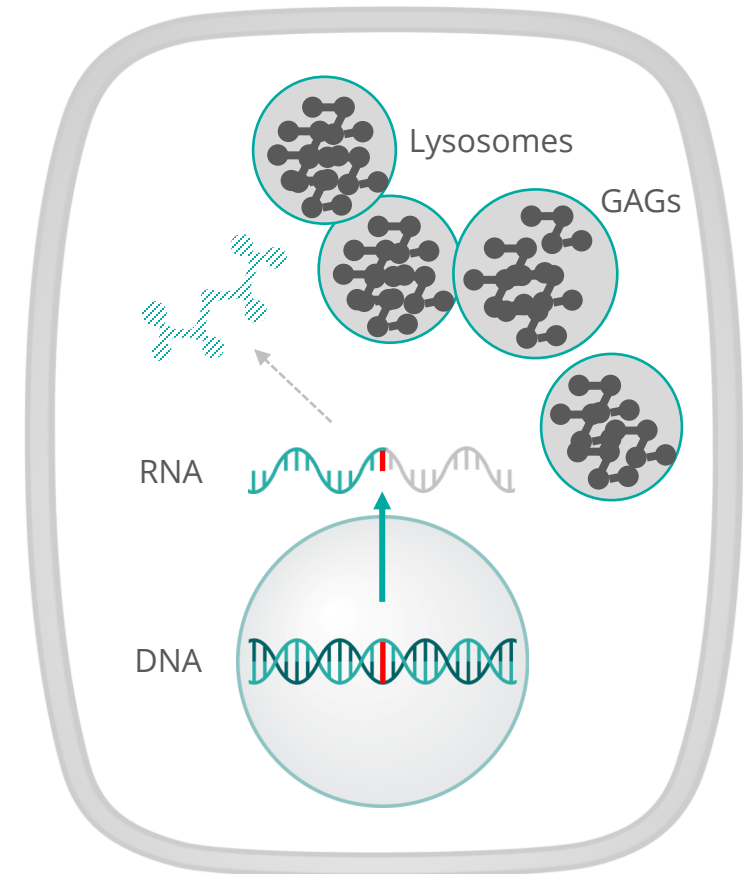
### Hurler syndrome

- Mucopolysaccharidosis type I
- Mutations in the *IDUA* gene
- Deficiency of the lysosomal **iduronidase** enzyme
- Accumulation of glycosaminoglycans (**GAGs**)
- *IDUA* W402X mutation most common cause: **UGG -> UAG**

WT *IDUA* gene



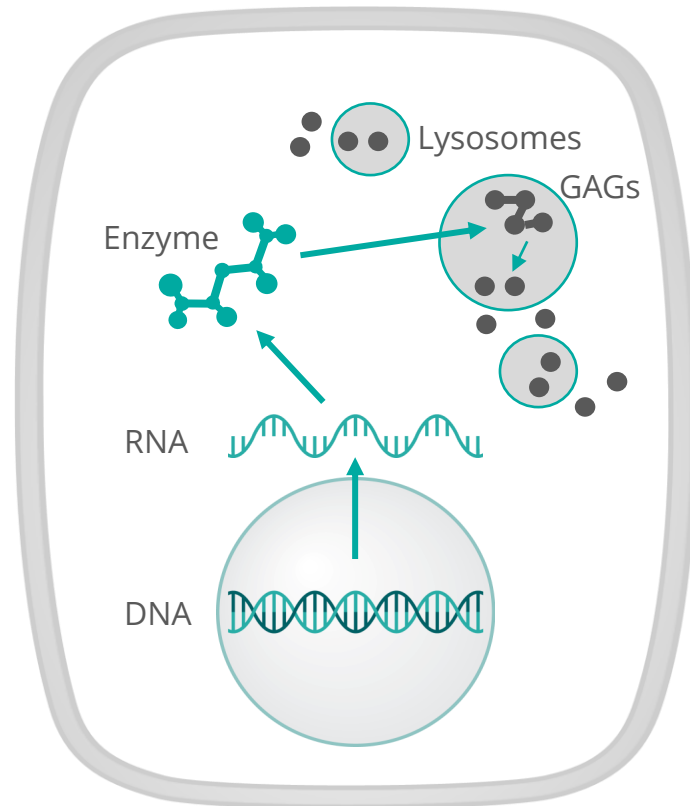
*IDUA* W402X mutant



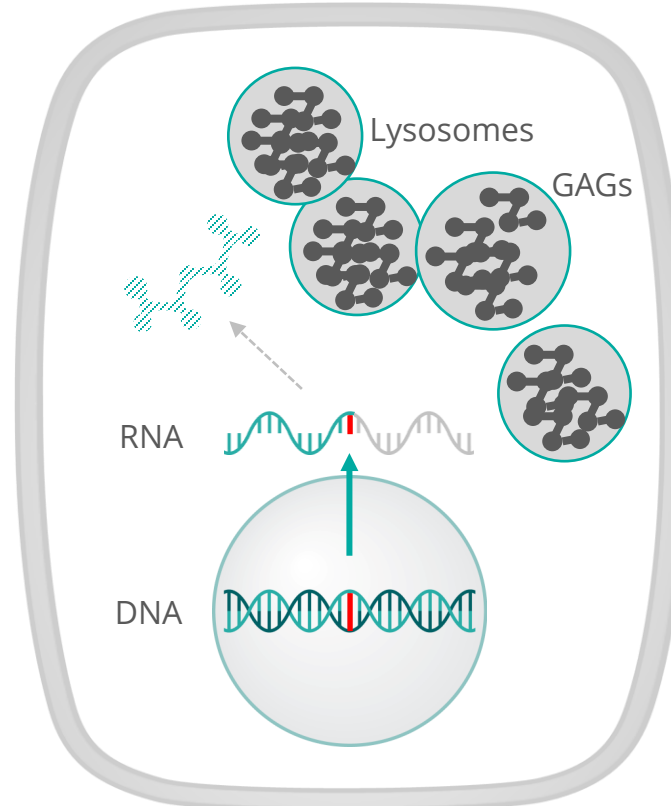
# Hurler syndrome – mouse model

Therapeutic approach using Axiomer® EONs

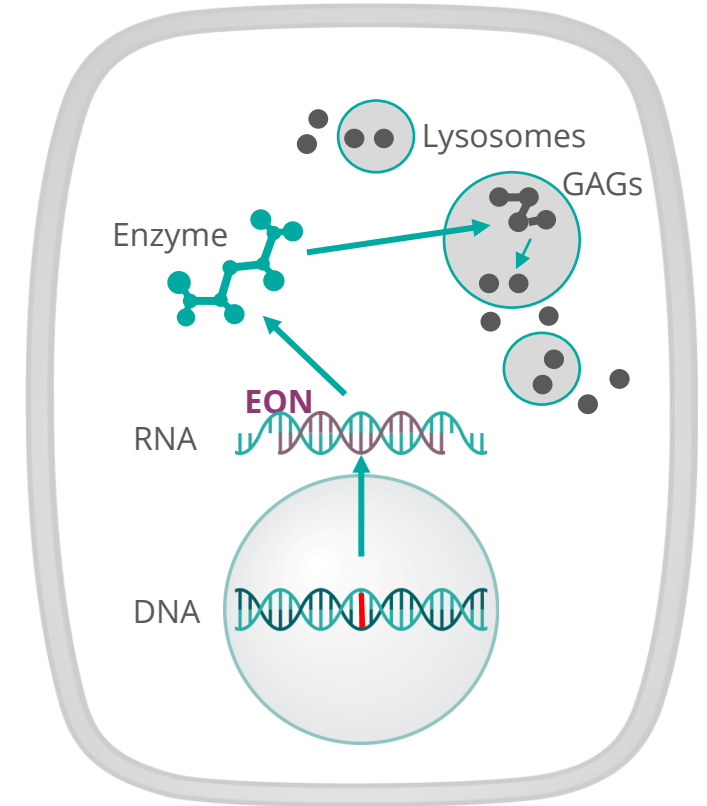
WT *Idua* gene



*Idua* W392X mutant

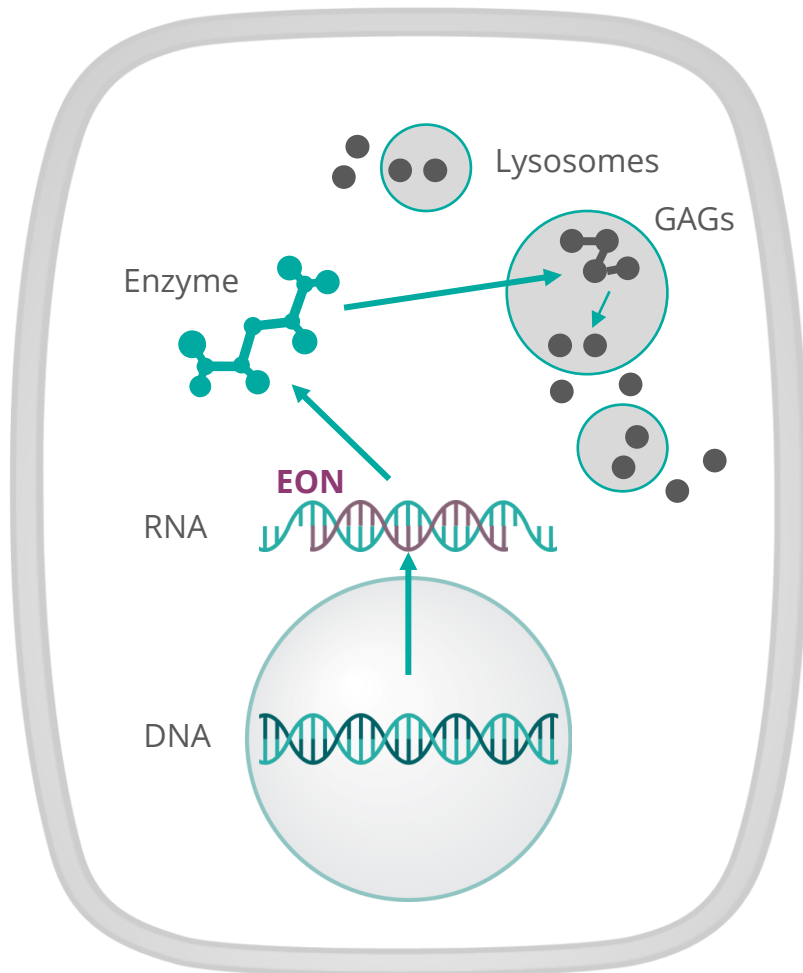


*Idua* W392X mutant and EON

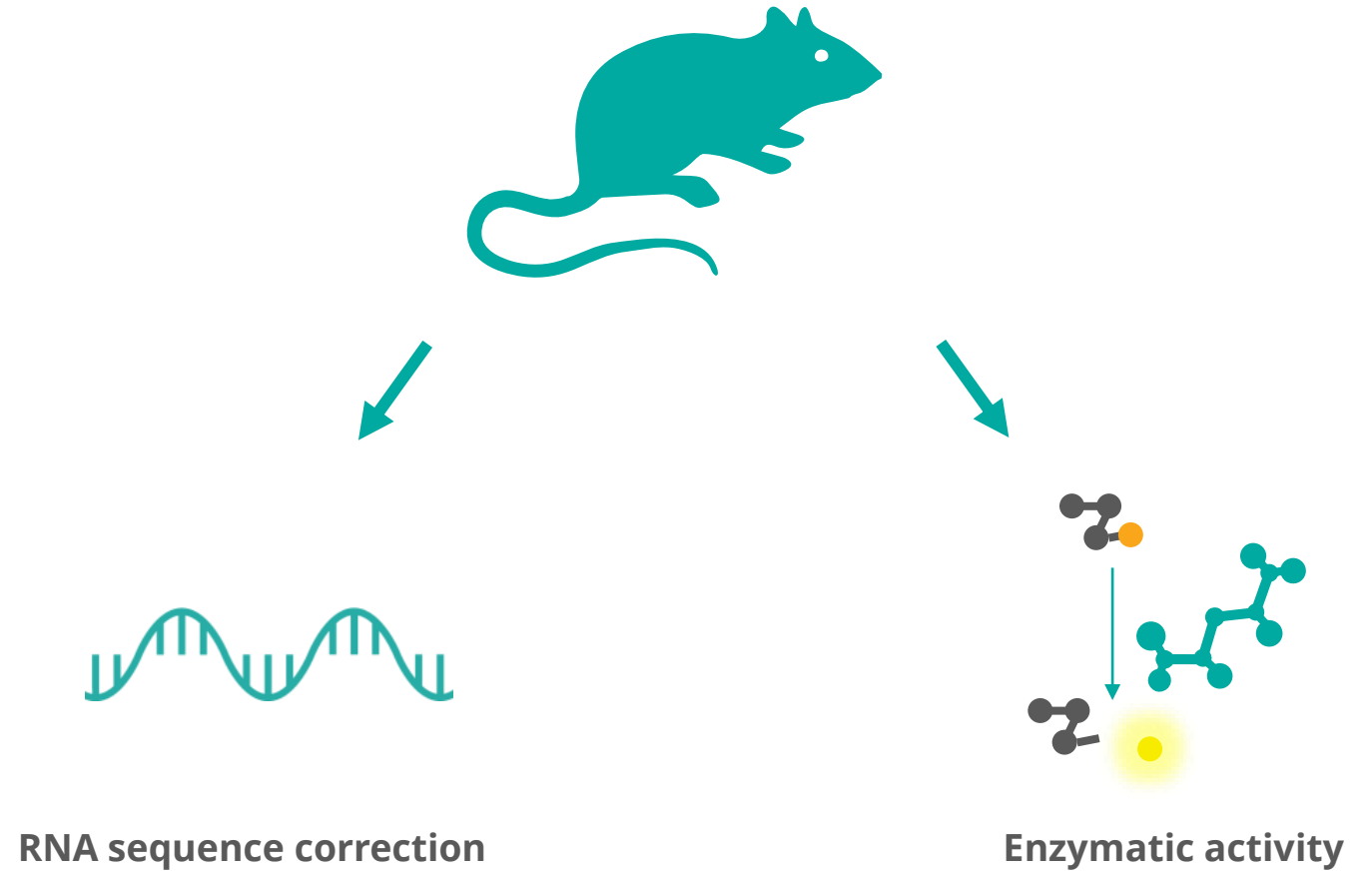


# Hurler mouse model for targeted editing

*Idua* W392X mutant and EON

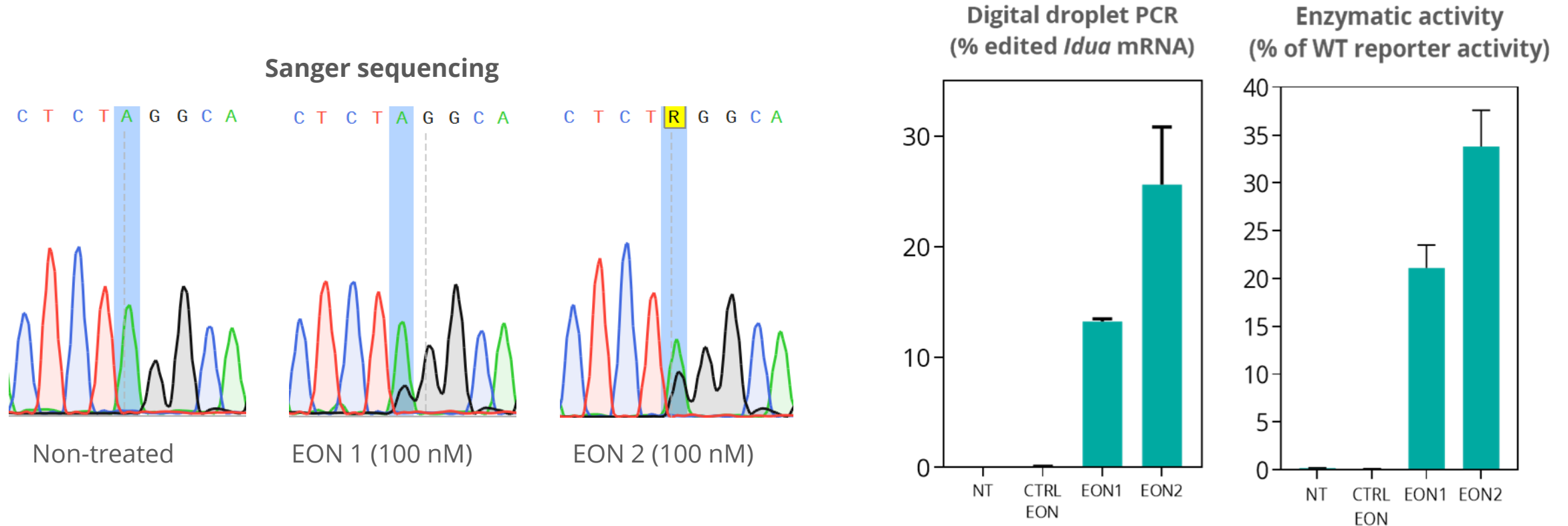


Readouts for restored function



# EONs correct *Idua* mRNA *in vitro*

Correction mediated by endogenous ADAR

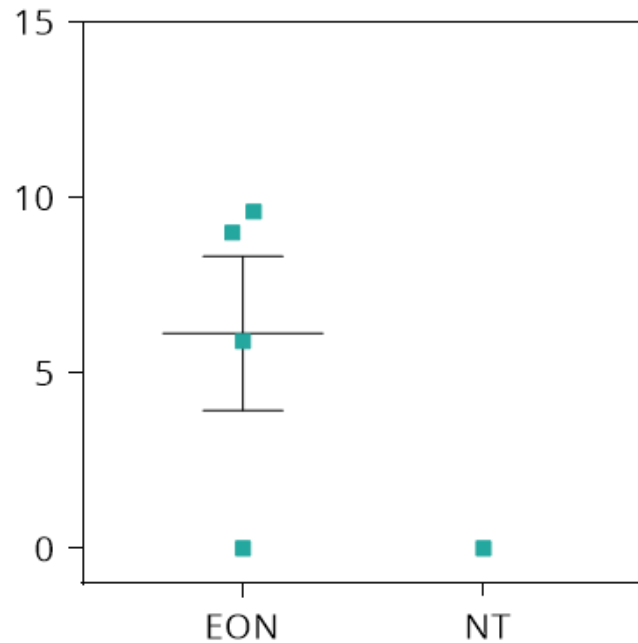


*Idua* W392X reporter construct in MEF cells with endogenous ADAR

# EONs correct *Idua* mRNA *in vivo*

Correction mediated by endogenous ADAR

Retina: % Edited *Idua* RNA



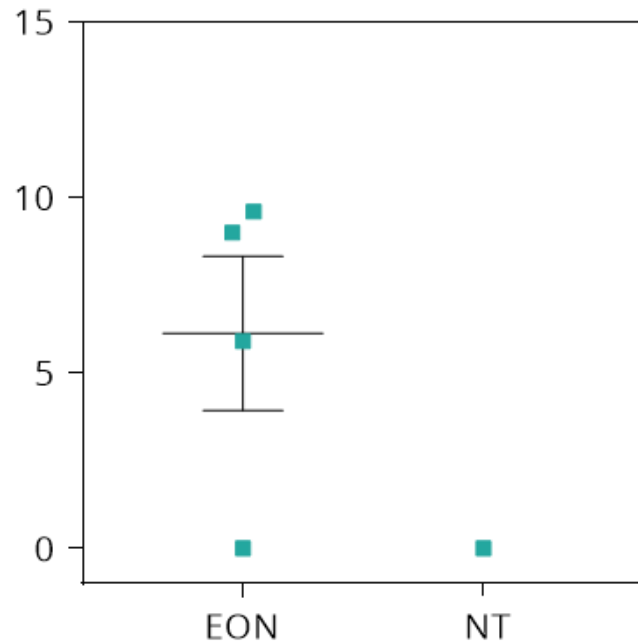
IVT injection of naked EON

Single dose 35  $\mu$ g EON; 7 days

# EONs correct *Idua* mRNA *in vivo*

*Efficacy similar to early phase exon skipping oligos*

Retina: % Edited *Idua* RNA



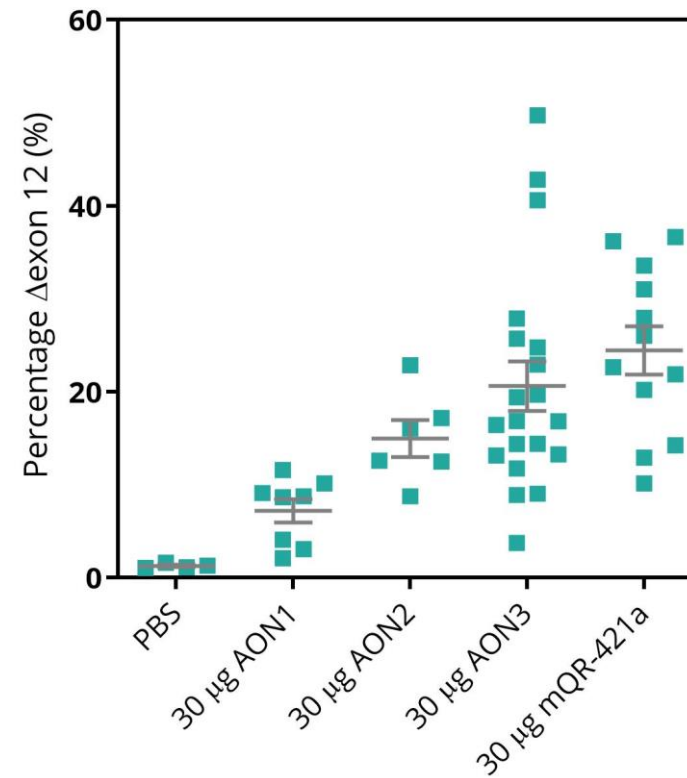
EONs are functional *in vivo* after single dose of 35  $\mu$ g. EONs are equally efficacious as early splice-switching oligos at comparable dose.

Further EON optimization ongoing.

IVT injection of naked EON

Single dose 35  $\mu$ g EON; 7 days

Retina: % Exon skip *Ush2a*

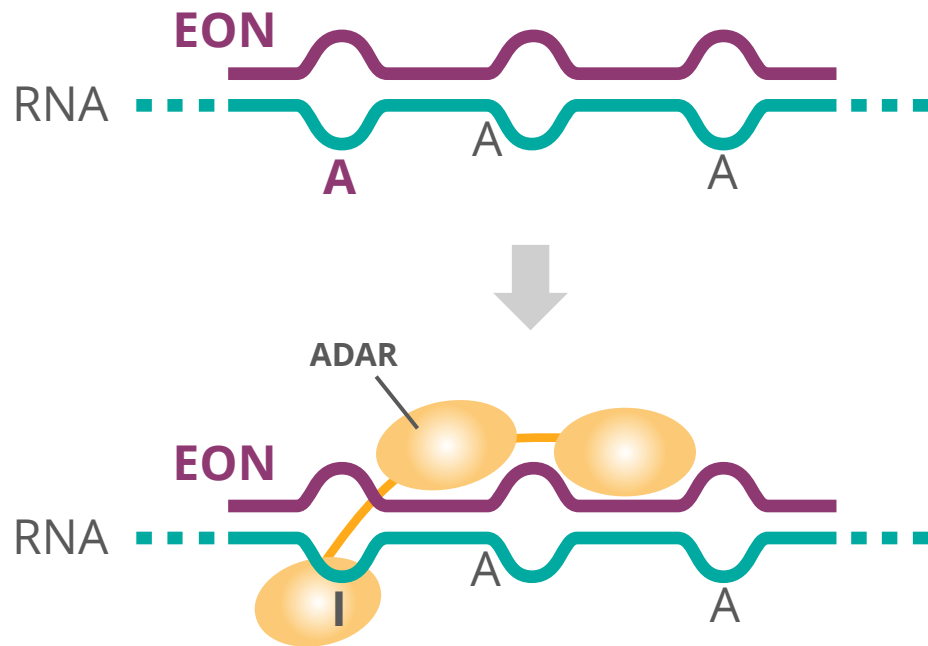


Splice-switching anti-sense oligos also needed optimization for improved skip levels.

QR-421a is now in clinical trials.



# Summary: Axiomer® technology



- Editing Oligonucleotides (EONs) recruit **endogenous** ADARs to catalyze **A-to-I** editing
- Editing occurs at **specific** adenosines of **endogenous** RNA transcripts
- Axiomer® is a **single-component** technology to **reversibly** modulate cellular functions
- Rational EON design enables optimization of editing **efficiency** and **drug-like** properties

The logo features the text "ProQR" in a white, bold, sans-serif font, centered within a white, hand-drawn, irregular oval shape. The background is a teal color with a repeating pattern of overlapping triangles in various shades of green and blue.

ProQR<sup>®</sup>